

**A Rare Case of Liddle Syndrome Due To A Novel Mutation in A Normotensive Adult**

<sup>1</sup>Dr Manjuri Sharma, Professor and HOD, Department of Nephrology, Gauhati Medical College and Hospital, Guwahati-32

<sup>2</sup>Dr Utpal Kalita, Resident, Department of Nephrology, Gauhati Medical College and Hospital, Guwahati-32

<sup>3</sup>Dr Prodip Kr Doley, Associate Professor, Department of Nephrology, Gauhati Medical College and Hospital, Guwahati-32

<sup>4</sup>Dr Gayatri Pegu, Associate Professor, Department of Nephrology, Gauhati Medical College and Hospital, Guwahati-32

<sup>5</sup>Dr Miranda Pegu, Associate Professor, Department of Nephrology, Gauhati Medical College and Hospital, Guwahati-32

<sup>6</sup>Dr Angelia L Khawbung, Resident, Department of Nephrology, Gauhati Medical College and Hospital, Guwahati-32

<sup>7</sup>Dr Nitin Dinesh, Resident, Department of Nephrology, Gauhati Medical College and Hospital, Guwahati-32

<sup>8</sup>Dr Moinul Islam, Resident, Department of Nephrology, Gauhati Medical College and Hospital, Guwahati-32

**Corresponding Author:** Dr Utpal Kalita, Resident, Department of Nephrology, Gauhati Medical College and Hospital, Guwahati-32

**How to citation this article:** Dr Manjuri Sharma, Dr Utpal Kalita, Dr Prodip Kr Doley, Dr Gayatri Pegu, Dr Miranda Pegu, Dr Angelia L Khawbung, Dr Nitin Dinesh, Dr Moinul Islam, “A Rare Case of Liddle Syndrome Due To A Novel Mutation in A Normotensive Adult”, IJMACR- June - 2025, Volume – 8, Issue - 3, P. No. 202 – 210.

**Open Access Article:** © 2025 Dr Utpal Kalita, et al. This is an open access journal and article distributed under the terms of the creative common's attribution license (<http://creativecommons.org/licenses/by/4.0>). Which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.

**Type of Publication:** Case Report

**Conflicts of Interest:** Nil

**Abstract**

Liddle syndrome (pseudoaldosteronism) is a genetically heterogenous autosomal dominant disorder. The key clinical characteristics of this syndrome are early onset salt-sensitive hypertension with hypokalemia, metabolic alkalosis, inhibition of renin activity and aldosterone secretion. Liddle syndrome is caused by mutations (missense or frameshift) in the genes of epithelial sodium channel (ENaC) subunits namely SCNN1A, SCNN1B or SCNN1G genes.

A normotensive adult diabetic male with CKD presenting with hypokalemic paralysis, was found to have Liddle syndrome after ruling out commoner causes. The genetic analysis revealed a novel mutation in  $\alpha$  subunit of ENaC. This is likely the second SCNN1A mutation identified after the description of one case of germ line mutation in the  $\alpha$  subunit in a Caucasian family affected by Liddle syndrome by Salih M.

**Keywords:** Liddle syndrome, Hyperkalemia, Hypertension, Chronic kidney disease, gene mutation

## Introduction

Liddle syndrome (pseudoaldosteronism) is a genetically heterogenous autosomal dominant disorder. The key clinical characteristics of this syndrome are early onset salt-sensitive hypertension with hypokalemia, metabolic alkalosis, inhibition of renin activity and aldosterone secretion<sup>1</sup>. Liddle syndrome is caused by mutations (missense or frameshift) in the genes of epithelial sodium channel (ENaC) subunits. ENaC channels are present in the apical portion of epithelial cells of distal nephron, distal colon, lung and ducts of exocrine glands. These channels belong to the ENaC/DEG (degenerin) family of proton-gated cation channels which mediates transport of Na<sup>+</sup> through the apical membrane from lumen into the epithelial cell<sup>2</sup>. By regulating Na<sup>+</sup> ion balance in the extracellular fluid (ECF) and in the kidney, these channels are a significant part of ECF volume and blood pressure (BP) regulation<sup>3</sup>.

## Case report

A 58-year-old male, diabetic for 4 years, normotensive, known chronic kidney disease (CKD) for 4 years, presented to our OPD with symptoms of weakness on the lower limbs. There was no history of leg swelling, urine output was adequate, glycemic control was Laboratory evaluation revealed the following

maintained. General examination did not reveal any positive finding except pallor. Neurological examination revealed diminished tone, diminished power (2/5) on both lower limbs with diminished deep tendon reflexes (flaccid paralysis). Other system examination was normal. Patient's BP was normal (126/70 mm Hg), no signs of peripheral edema. The patient was on oral antidiabetic drug linagliptin.

He didn't give any history of fever, diarrhea, vomiting, excessive urine volume or of any diuretic intake.

Routine testing revealed hypokalemia (K<sup>+</sup> 2.6 mmol/dl), S.creatinine 3.2 mg/dl, Hemoglobin 8.8 g/dl. The rest of the parameters were under normal limits. He was prescribed oral potassium supplementation and previous medications were continued. However, the serum potassium was persistently low despite intravenous correction. Further evaluation was planned, a battery of tests advised which is as follows.

Meanwhile, a Neurology consultation was taken, and the patient also underwent a nerve conduction study (NCS) to rule out peripheral neuropathy (diabetic neuropathy). The study revealed diminished cMAP with normal conduction velocities. There was no clinical or radiological evidence of myelopathy.

Parameter	Observed value	Normal range
Blood parameters		
WBC	8600	4.00-11.00 x 10 <sup>3</sup> /μL
Hemoglobin	9.4	13-16.5 g/dl
Platelet count	231,000	150-400 x 10 <sup>3</sup> / μL
RBS	146	74.0-106.0 mg/dl
Urea	70	14.98-36.38 mg/dl
Creatinine	3.4	0.52-1.05 mg/dl
Na	137	137-145 mmol/l
K	1.8	3.5-5.1 mmol/l

Ca	9.3	8.40-10.20 mg/dl
Mg	1.5	1.6-2.3 mg/dl
PO4	4.2	2.50-4.50 mg/dl
Uric acid	5.8	2.50-8.50 mg/dl
Albumin	4.5	3.5-5.0 g/dl
TSH	0.79	0.40-4.05 mIU/L
Urine routine		
Albumin	Trace	Negative
Glucose	Nil	Negative
Pus cells	5-6/hpf	1-5/hpf
Epithelial cells	2-3/hpf	Upto 20/hpf
RBC	Nil	Absent
Ultrasound Abdomen		
Bilaterally contracted kidneys RK - 7.8 cm, LK – 8.4 cm, corticomedullary differentiation blurred. Rest organs within normal limits.		
Urine electrolytes		
Na	58 mmol/L	
K	23.7 mmol/L	
Ca	3.7 mg/dl	
Cl	82 mmol/L	
ABG		
pH	7.57	7.35-7.45
pCO2	39.6	36.0-48.0 mmHg
HCO3	29.3	22-26 mmol/L

Repeated testing of serum electrolytes revealed persistent hypokalemia. A thorough drug history was taken and a review of medication which might lead to hypokalemia was done and ruled out. The thyroid function test was normal. ABG was suggestive of metabolic alkalosis.

To rule out urinary potassium loss, urine electrolytes were tested which showed higher than normal potassium excretion. Calculated TTKG was found to be 5.

Based on ruling out other causes of hypokalemia, a presumptive clinical diagnosis of a potassium-losing

channelopathy was made. As the patient had hypokalemic metabolic alkalosis without hypertension, we suspected either Bartter or Gitelman syndrome. Further considering the age of the patient and low urinary Ca and concomitant hypomagnesemia, a diagnosis of Gitelman syndrome was more likely.

To confirm our case, we sent a gene panel test for hypokalemic disorders, which revealed, contrary to our clinical diagnosis, a likely pathogenic variant mutation of ENaC channel leading to Liddle syndrome.

Parents are non-consanguineously married.  
There is no family history of similar complaints.  
**RESULT SUMMARY**

**Positive – Likely Pathogenic variant detected**

#### VARIANT DETAILS

GENE/REFSEQ	COORDINATE (GRCh37)	VARIANT* (107s)	EXON/INTRON	VARIANT TYPE	ZYGOSITY/INHERITANCE	OMIM/PHENOTYPE	CLASSIFICATION* ACMG/AMP
SCNN1A	chr12:6483916	c.32_33insT p.Ser125*1	Exon 2	Frameshift	Heterozygous/AD	Liddle syndrome-3 (OMIM#618126)	Likely Pathogenic

\*Clinical correlation Recommended

#### IN SILICO PREDICTION PARAMETERS

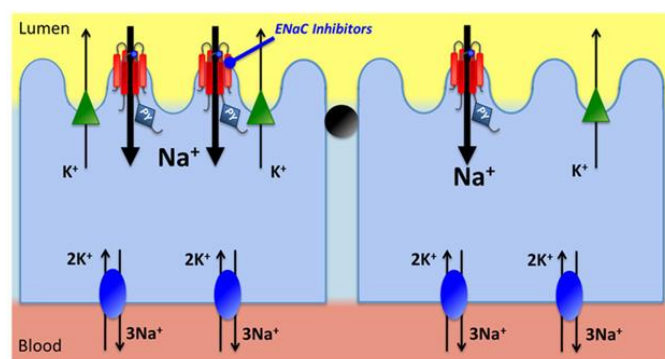
Variant	EFFECT
MutationTaster	Damaging

The patient was put on oral amiloride tablet 5 mg along with oral potassium supplements. Subsequently, potassium levels increased to normal range and the patient improved clinically with improvement in muscle power. Within a few days, the patient was off potassium supplements and maintained normal electrolytes with amiloride alone. The patient is doing well till now.

## Discussion

ENaC is an amiloride-sensitive epithelial sodium channel, localized in the apical portion of epithelial cells of distal nephron<sup>4</sup>. Under physiological conditions, its expression and activity are positively regulated by aldosterone and antidiuretic hormone and are influenced by numerous extracellular factors, such as sodium, chloride, protons and proteases<sup>5</sup>. This channel is crucial, together with ROMK (renal outer medullary K<sup>+</sup>) channels and Na<sup>+</sup>/K<sup>+</sup> ATPase, for Na<sup>+</sup> reabsorption and K<sup>+</sup> excretion and thus, for electrolytes homeostasis<sup>5</sup>. The channel is a heteromeric complex constituted of three homologous subunits,  $\alpha$ ,  $\beta$ , and  $\gamma$ <sup>4,6,7</sup>, encoded by the SCNN1A (chr 12p13.31), SCNN1B (chr 16p12.2) and SCNN1G (chr 16p12.2) genes, respectively<sup>5</sup>. Within the C-terminus of all three ENaC subunits, there is a highly conserved sequence, named the PY (Proline Tyrosine) motif, which is a binding site for Nedd4 (Neural precursor cell expressed, developmentally down-regulated 4), that mediates internalization and proteasomal degradation of the channel<sup>5,8-10</sup>.

Liddle syndrome results from germline mutations in SCNN1A, SCNN1B or SCNN1G genes. The germline mutations of the SCNN1B and SCNN1G genes cause loss or disruption of the PY sequence, resulting in gain-of-function and determine increased membrane density of ENaC and consequent increase in renal Na<sup>+</sup> reabsorption. The germline mutation of SCNN1A gene affects the extracellular domain, causing disruption of a disulfide bridge, leading to gain-of-function and increase of the open probability of the channel and consequent increase in Na<sup>+</sup> current, without affecting the PY motif.



The prevalence of Liddle syndrome across the general hypertensive population is unknown. In two recent studies, including 330 and 766 Chinese patients affected by arterial hypertension, the prevalence of Liddle syndrome resulted to be 1.52%<sup>11</sup> and 0.91%<sup>12</sup>, respectively.

The typical clinical feature is resistant, early onset salt-sensitive hypertension, often associated with a family history of early onset hypertension and sudden death. Biochemically, the characteristic findings are hypokalemia, metabolic alkalosis, suppressed plasma renin activity (PRA) and low serum aldosterone levels.

## Crosstalk between diabetes and ENaC:

Diabetes mellitus is the leading cause of a form of chronic kidney disease known as Diabetic Kidney Disease (DKD), which is the most common cause of end-stage renal disease (ESRD). DKD is associated with

significant changes in renal hemodynamics and electrolyte transport. Alterations in renal ion transport, triggered by pathophysiological conditions in diabetes, can exacerbate hypertension, accelerate renal injury, and are integral to the development of DKD. Some of the major renal ion transporters have altered expression and activity under diabetic conditions including SGLT2, ENaC, TRPC6, TRPM6, NHE,  $K_{ATP}$ .

### ENaC activity in diabetes

Expressed primarily in principal cells of the distal nephron, ENaC plays a central role in maintaining salt and water homeostasis, regulating extracellular fluid volume, controlling blood pressure, and overall renal function<sup>5</sup>. Diabetes and DKD have been associated with increased ENaC activity and expression by multiple mechanisms, which may interfere with renal blood pressure control, exacerbate hypertension, and thereby contribute to the progression of DKD.

Studies have shown that insulin augments ENaC expression and activity<sup>15,16,17</sup>. As an example, single-channel analysis in freshly isolated, split-open tubules demonstrated that ENaC activity was acutely activated by insulin. Moreover, insulin receptor knockout mice have significantly lower activity compared to their wild-type littermates<sup>18</sup>.

One proposed mechanism suggests that ENaC involvement with DKD is inexorably linked to the serum and glucocorticoid-regulated kinase (SGK1) protein. SGK1 is stimulated by insulin, which causes more ENaC to be translocated to the membrane (through the Nedd4–2 signaling pathway) increasing sodium reabsorption from the tubule. This may lead to excess renal sodium retention, hypertension, and ultimately renal damage associated with DKD<sup>19</sup>. In a rat model of streptozotocin (STZ)-induced T1DM, increased glucose was correlated

with upregulation of all three ENaC subunits, attributed to elevations in aldosterone and vasopressin<sup>20</sup>. Another proposed mechanism of ENaC increases in DKD involves the serine protease, plasmin<sup>21</sup>. Dysfunction of the glomerular filtration barrier (GFB) in DKD causes plasmin to be filtered to the tubules where it activates ENaC and increases sodium reabsorption<sup>22</sup>. In a study of patients with T2DM, microalbuminuria, a hallmark of GFB breakdown, is associated with increased aberrant filtration of plasmin. This surge of filtered plasmin was shown to be sufficient to increase the open probability for ENaC and was proposed as a possible mechanism contributing to hypertension in diabetes<sup>23</sup>. Clinical studies have also found that amiloride, an ENaC blocker, may be protective in DKD as it significantly increased sodium excretion, and reduced blood pressure, albuminuria, and plasmin in urine of diabetic patients<sup>24</sup>. Recently a pilot randomized cross-over study comparing the effects of daily administration of either oral amiloride or the NCC inhibitor, hydrochlorothiazide (HCTZ), to patients with type 2 diabetes and proteinuria revealed similar effects with both drugs resulting in reduced systolic blood pressure<sup>25</sup>.

In diabetes, prolonged hyperglycemia causes excess glucose to contact and react with proteins and lipids resulting in advanced glycation end-products (AGEs), with an especially pronounced elevation in the distal nephron where ENaC is highly expressed<sup>26</sup>. When applied to cultured tubular epithelial cells in concentrations comparable to what occurs in diabetes, AGEs increased ENaC mRNA and protein and stimulated ENaC activity by inhibiting catalase and increasing intracellular ROS production<sup>27</sup>. The effect on ENaC activity persisted for more than 72 hours after removal of AGEs. This sustained ENaC elevation may

be key to understanding why DKD often continues to progress despite adequate glucose control and provide key insights necessary for the development of more effective treatments.

From these studies, it is evident that diabetes creates pathophysiological conditions that affect ENaC via multiple pathways, causing a sustained increase in activity or expression, ultimately resulting in blood pressure elevation. As hypertension is one of the most important risk factors in the progression from diabetes to DKD, ENaC is a critical mechanistic and potential therapeutic target in DKD research.

### **Liddle syndrome and CKD**

One of the hallmarks of Liddle syndrome is elevated blood pressure due to the overactive sodium channels in the kidneys. Chronic hypertension is a well-known risk factor for the development of CKD (hypertensive nephrosclerosis). Another mechanism of kidney damage in Liddle syndrome is prolonged hypokalemia. Over time, the chronic effects of high blood pressure, electrolyte imbalances, and kidney damage may contribute to kidney dysfunction, which could potentially progress to CKD.

### **Our case of Liddle syndrome**

The patient of Liddle syndrome presented here is unique because unlike the most common presentation of early-onset resistant hypertension, our patient had a normal BP. It is seen that extremely severe phenotypes and mild forms can exist, with some patients carrying a causative mutation who are normotensive or in whom a clinical diagnosis of Liddle syndrome is made in old age<sup>14</sup>. Moreover, in this case there was no family history of young-onset hypertension or known monogenic disease. The only clue to a genetic basis of the disease was

hypokalemic metabolic alkalosis, for which no known factor could be found.

Although the patient was diabetic, it was relatively short duration, and the glycemic control was maintained. Moreover, there were no clinical features of diabetic kidney disease like edema, visual disturbance etc, and paraparesis could not be attributed to diabetic neuropathy as tested by NCS study. A fundus examination was performed, and diabetic retinopathy was ruled out. To clinch the exact diagnosis of kidney dysfunction, we had planned for a renal biopsy but could not go ahead due to bilaterally small sized kidneys with altered cortico-medullary differentiation (CMD).

Based on these observations, we can presume that the chronic kidney dysfunction could be attributed to Liddle syndrome and associated dyselectrolytemia (chronic hypokalemia).

Another unique feature is the mutation described in the genetic analysis report. It is a novel heterozygous frameshift mutation c.32\_33insT p.Ser12fs\*1 in chr12 : 6483916 exon 2 of the SCNN1A (NM\_001038.6) by NGS analysis. According to ACMG recommendation, this mutation is classified as likely pathogenic. This is a very rare entity, and this particular mutation has never been described before in literature.

Of all reported cases of Liddle syndrome, till date 24 different mutations of the  $\beta$  subunit and 6 different mutations of the  $\gamma$  subunit were identified in 72 families worldwide. Only one case of germ line mutation in the  $\alpha$  subunit was identified in a Caucasian family affected by Liddle syndrome. This is likely the second mutation identified in  $\alpha$  subunit of ENaC<sup>13</sup>.

### **Conclusion**

This is the first case of Liddle syndrome reported by our center. A normotensive adult diabetic male with CKD



presenting with hypokalemic paralysis, was found to have Liddle syndrome after ruling out commoner causes. Our results expand the mutational spectrum of Liddle syndrome, this likely being the second  $\alpha$ -subunit mutation to be reported in literature. The rare syndromes, owing to variants of the same gene mutation, may have varied clinical presentation, therefore it is utmost important to have a high degree of suspicion for these uncommon diagnoses. The role of genetic testing in appropriate clinical setting cannot be overemphasized.

## References

1. Yang KQ, Xiao Y, Tian T, Gao LG, Zhou XL. Molecular genetics of Liddle's syndrome. *Clinica Chimica Acta*. 2014 Sep 25;436:202-6.
2. Garty H, Palmer LG. Epithelial sodium channels: function, structure, and regulation. *Physiological reviews*. 1997 Apr 1;77(2):359-96.
3. Büsst CJ. Blood pressure regulation via the epithelial sodium channel: from gene to kidney and beyond. *Clinical and Experimental Pharmacology and Physiology*. 2013 Aug;40(8):495-503.
4. Canessa CM, Schild L, Buell G, Thorens B, Gautschi I, Horisberger JD, Rossier BC. Amiloride-sensitive epithelial Na<sup>+</sup> channel is made of three homologous subunits. *Nature*. 1994 Feb 3;367(6462):463-7.
5. Hanukoglu I, Hanukoglu A. Epithelial sodium channel (ENaC) family: Phylogeny, structure–function, tissue distribution, and associated inherited diseases. *Gene*. 2016 Apr 1;579(2):95-132.
6. Staruschenko A, Medina JL, Patel P, Shapiro MS, Booth RE, Stockand JD. Fluorescence resonance energy transfer analysis of subunit stoichiometry of the epithelial Na<sup>+</sup> channel. *Journal of Biological Chemistry*. 2004 Jun 25;279(26):27729-34.
7. Jasti J, Furukawa H, Gonzales EB, Gouaux E. Structure of acid-sensing ion channel 1 at 1.9 Å resolution and low pH. *Nature*. 2007 Sep 20;449(7160):316-23.
8. Schild L, Lu Y, Gautschi I, Schneeberger E, Lifton RP, Rossier BC. Identification of a PY motif in the epithelial Na channel subunits as a target sequence for mutations causing channel activation found in Liddle syndrome. *The EMBO journal*. 1996 May 15;15(10):2381-7.
9. Rotin D, Staub O. Role of the ubiquitin system in regulating ion transport. *Pflügers Archiv-European Journal of Physiology*. 2011 Jan;461:1-21.
10. Butterworth MB. Regulation of the epithelial sodium channel (ENaC) by membrane trafficking. *Biochimica Et Biophysica Acta (BBA)-Molecular Basis of Disease*. 2010 Dec 1;1802(12):1166-77.
11. Wang LP, Yang KQ, Jiang XJ, Wu HY, Zhang HM, Zou YB, Song L, Bian J, Hui RT, Liu YX, Zhou XL. Prevalence of Liddle syndrome among young hypertension patients of undetermined cause in a Chinese population. *The Journal of Clinical Hypertension*. 2015 Nov;17(11):902-7.
12. Liu K, Qin F, Sun X, Zhang Y, Wang J, Wu Y, Ma W, Wang W, Wu X, Qin Y, Zhang H. Analysis of the genes involved in Mendelian forms of low-renin hypertension in Chinese early-onset hypertensive patients. *Journal of hypertension*. 2018 Mar 1;36(3):502-9.
13. Salih M, Gautschi I, Van Bemmelen MX, Di Benedetto M, Brooks AS, Lugtenberg D, Schild L, Hoorn EJ. A missense mutation in the extracellular domain of  $\alpha$ ENaC causes Liddle syndrome. *Journal of the American Society of Nephrology*. 2017 Nov 1;28(11):3291-9.

14. Pepersack T, Allegre S, Jeunemaître X, Leeman M, Praet JP. Liddle syndrome phenotype in an octogenarian. *The Journal of Clinical Hypertension*. 2014 Nov 27;17(1):59.
15. Gonzalez-Rodriguez E, Gaeggeler HP, Rossier BC. IGF-1 vs insulin: respective roles in modulating sodium transport via the PI-3 kinase/Sgk1 pathway in a cortical collecting duct cell line. *Kidney international*. 2007 Jan 2;71(2):116-25.
16. Ilatovskaya DV, Levchenko V, Brands MW, Pavlov TS, Staruschenko A. Cross-talk between insulin and IGF-1 receptors in the cortical collecting duct principal cells: implication for ENaC-mediated Na<sup>+</sup> reabsorption. *American Journal of Physiology-Renal Physiology*. 2015 Apr 1;308(7):F713-9.
17. Mansley MK, Watt GB, Francis SL, Walker DJ, Land SC, Bailey MA, Wilson SM. Dexamethasone and insulin activate serum and glucocorticoid-inducible kinase 1 (SGK1) via different molecular mechanisms in cortical collecting duct cells. *Physiological reports*. 2016 May;4(10):e12792.
18. Pavlov TS, Ilatovskaya DV, Levchenko V, Li L, Ecelbarger CM, & Staruschenko A (2013). Regulation of ENaC in mice lacking renal insulin receptors in the collecting duct. *FASEB J*, 27(7), 2723–2732. 10.1096/fj.12-223792 [PubMed: 23558339]
19. McCormick JA, Bhalla V, Pao AC, Pearce D. SGK1: a rapid aldosterone-induced regulator of renal sodium reabsorption. *Physiology*. 2005 Apr;20(2):134-9.
20. Song J, Knepper MA, Verbalis JG, Ecelbarger CA. Increased renal ENaC subunit and sodium transporter abundances in streptozotocin-induced type 1 diabetes. *American Journal of Physiology-Renal Physiology*. 2003 Dec;285(6):F1125-37.
21. Ray EC, Miller RG, Demko JE, Costacou T, Kinlough CL, Demko CL, Unruh ML, Orchard TJ, Kleyman TR. Urinary plasmin (ogen) as a prognostic factor for hypertension. *Kidney international reports*. 2018 Nov 1;3(6):1434-42.
22. Svenningsen P, Skøtt O, Jensen BL. Proteinuric diseases with sodium retention: is plasmin the link?. *Clinical and Experimental Pharmacology and Physiology*. 2012 Jan;39(1):117-24.
23. Buhl KB, Oxlund CS, Friis UG, Svenningsen P, Bistrup C, Jacobsen IA, Jensen BL. Plasmin in urine from patients with type 2 diabetes and treatment-resistant hypertension activates ENaC in vitro. *Journal of hypertension*. 2014 Aug 1;32(8):1672-7.
24. Andersen H, Friis UG, Hansen PB, Svenningsen P, Henriksen JE, Jensen BL. Diabetic nephropathy is associated with increased urine excretion of proteases plasmin, prothrombin and urokinase and activation of amiloride-sensitive current in collecting duct cells. *Nephrology Dialysis Transplantation*. 2015 May 1;30(5):781-9.
25. Unruh ML, Pankratz VS, Demko JE, Ray EC, Hughey RP, Kleyman TR. Trial of amiloride in type 2 diabetes with proteinuria. *Kidney international reports*. 2017 Sep 1;2(5):893-904.
26. Schleicher ED, Wagner E, Nerlich AG. Increased accumulation of the glycoxidation product N (epsilon)-(carboxymethyl) lysine in human tissues in diabetes and aging. *The Journal of clinical investigation*. 1997 Feb 1;99(3):457-68.
27. Wang Q, Song B, Jiang S, Liang C, Chen X, Shi J, Li X, Sun Y, Wu M, Zhao D, Zhang ZR. Hydrogen Sulfide Prevents Advanced Glycation End-Products



Induced Activation of the Epithelial Sodium  
Channel. Oxidative Medicine and Cellular  
Longevity. 2015;2015(1):976848.